



2018/19 Project & 2020/21 Plan

ASTA G X E Cooperators' Meeting

December 5th, 2017

Project Concept for 2018-19 Testing Years:

- ❖ Sample alleles for G X E from ex-PVP, old, and unselected pool
- ❖ Dissect Good G X E (mechanisms allowing specific adaptation) vs Bad G X E (poor performance in specific environments due to poor alleles such as disease susceptibility)

Parameters:

- ❖ Substrate for:
 - ❖ Phenotyping
 - ❖ Modeling/Genome Prediction
- ❖ Use materials with a relatively narrow maturity window
 - ❖ Reduce impact of flowering time on results
 - ❖ More uniform data
 - ❖ Simplify plot management for collaborators
- ❖ Use same genetics across all locations
 - ❖ Division of 2016-17 experiments within G X E is not fully leveraging the breadth of environments



Project Plan:

- ❖ Populations - mini-NAM DH:
 - ❖ PHW65 X PHN11 (ex-PVP x ex-PVP) – “Good” allele source
 - ❖ PHW65 X Mo44 (ex-PVP x Old) - “Bad” allele source
 - ❖ PHW65 X MoG (ex-PVP x Unselected) – “Ugly” allele source
- ❖ Tester:
 - ❖ LH195 (B37 X B73) Midwest to South
 - ❖ PHT69 (B14 X B37) for North
- ❖ Yellow Stripe hybrids (30 plots/location)
 - ❖ Accumulate G X Y data and link previous work to new locations
- ❖ Continue a small component of the ex-PVP Design II hybrids or TX714 vs B73 and NILAS testing (~100 plots at a subset of locations)



Things to Still Think About:

- ❖ Allocation of PHT69 vs LH195 hybrids
- ❖ Proportion (10 – 20%) of hybrids replicated at each site
 - ❖ Possibly those hybrids already included in the 2014 & 15 experiment to allow calculation of G X Y estimates
 - ❖ Same hybrids replicated across locations
- ❖ Design?
 - ❖ Randomizing across populations allow direct comparison or block by population?
- ❖ Phenotyping
 - ❖ Can we implement phenotyping technologies across sites – EarthSense rover
- ❖ Mini-weather stations within canopy
 - ❖ Interest in the data?

Parameters for the G2F G X E Scientific Project - 2020/21 and Beyond Plan:

- ❖ Research should be founded on relevant research questions
- ❖ Materials (germplasm) used should be publicly available
- ❖ We need to be able to generate hybrid seed for (at least) 500 plots across (at least) 35 locations within the needed timeframe
- ❖ Generate good experiments: minimize the potential for external confounding factors on performance (experiments that fall apart, maturity, tester, etc)



Proposal for G2F G X E Science Project & Timeline:

- ❖ Can we characterize Midwest alleles well enough to determine the value of exotic alleles for productivity
- ❖ The idea is that molecular information allows us to determine what is different but not necessarily what is useful
- ❖ Immediately:



- ❖ Longer term: Develop a germplasm generation infrastructure that provides useful resources to the community over time

Proposed Plan: North America Germplasm-Based Genetic Modelling and Allele Characterization

Core Inbred pool
ex-PVP and public breeding inbreds

Stiff Stalk
- ex-PVP
- Public

Non-Stiff Stalk
- ex-PVP
- Public

Iodent
- ex-PVP
- Public

New Allele Sources

- GEM selects
- Particular lines from cooperators (yield component traits)
- Donations from industry screening
- Other sources

- Model per se allelic effects
 - Prediction of breeding crosses
 - Prediction of “ideal” inbreds
- Hybrid Prediction
- Traits of focus
- GxE
- Integrating phenotyping, etc.

- Determine how to characterize alleles in the context of Core Inbred Pool
 - Genetic context
 - Recombination/resolution
 - Managed biotic and abiotic stress targets

A Start:

STIFF STALK SET	PHBW8 PHJ40 type	3AZA1 B14 type	PHB47 B37 type	FBLL B73 type
GEMS-0061 <i>Cristalino Colorado</i>	DH1	DH2	DH3	DH4
GEMS-0113 <i>Tuxpeño</i>	DH5	DH6	DH7	DH8
GEMS-0219 <i>GEM x GEM derived</i>	DH9	DH10	DH11	DH12
GEMS-0227 <i>Inbred-tropical</i>	DH13	DH14	DH15	DH16

NON STIFF STALK SET	PHP02 <i>Iodent</i> type	PHJ89 PHT77 X PHG47	PHN46 PH848/814 /595 type	LH212Ht Mo17 type	PHK76 PHK76 type
GEMN-0096 <i>Hybrid-tropical</i>	DH17	DH18	DH19	DH20	DH21
GEMN-0097 <i>Mixed</i>	DH22	DH23	DH24	DH25	DH26
GEMN-0192 <i>Tusón</i>	DH27	DH28	DH29	DH30	DH31
GEMN-0225 <i>Composite (Suwan)</i>	DH32	DH33	DH34	DH35	DH36

Other Things to Consider:

- ❖ Is there value on evaluating similar sets of materials across environments
- ❖ Utility of managed locations to identify extreme genotypes for testing
- ❖ Deploying common technologies across sites

Deliverables:

- ❖ 2014 data publication - Nature Communications 8: 1348 (2017)
- ❖ Summary of 2014-15 data – Diego Jarquin & Aaron Lorenz
- ❖ Others:
 - ❖ Inbred evaluation - Celeste Falcon
 - ❖ 2014-2016/7 – hybrid combined analysis – Jim Holland & Anna Rogers
 - ❖ Maize Atlas NILAS project – Randy Wisser
 - ❖ NIL data – Zhi Li, Candy Hirsch & Nathan Springer
 - ❖ BSSS experiment – Bridget McFarland
 - ❖ Stalk lodging GWAS – Alex Lipka
 - ❖ "Yellow Stripe" manuscript – Liz Lee
 - ❖ Diallel set – Jode Edwards, Liz Lee, Martin Bohn

Field Season	Release to Collaborators (ARK)	Release to Public (DOI or ARK)
2014		http://dx.doi.org/10.7946/P2201Q
2015	February 2017	https://doi.org/10.7946/P24S31
2016	March 2017	ARK
N	March of N+1	March of N+2
Ears Imaging		Through Cyverse

Genomes To Fields Sponsors



United States Department of Agriculture
National Institute of Food and Agriculture



Genomes To Fields Collaborators

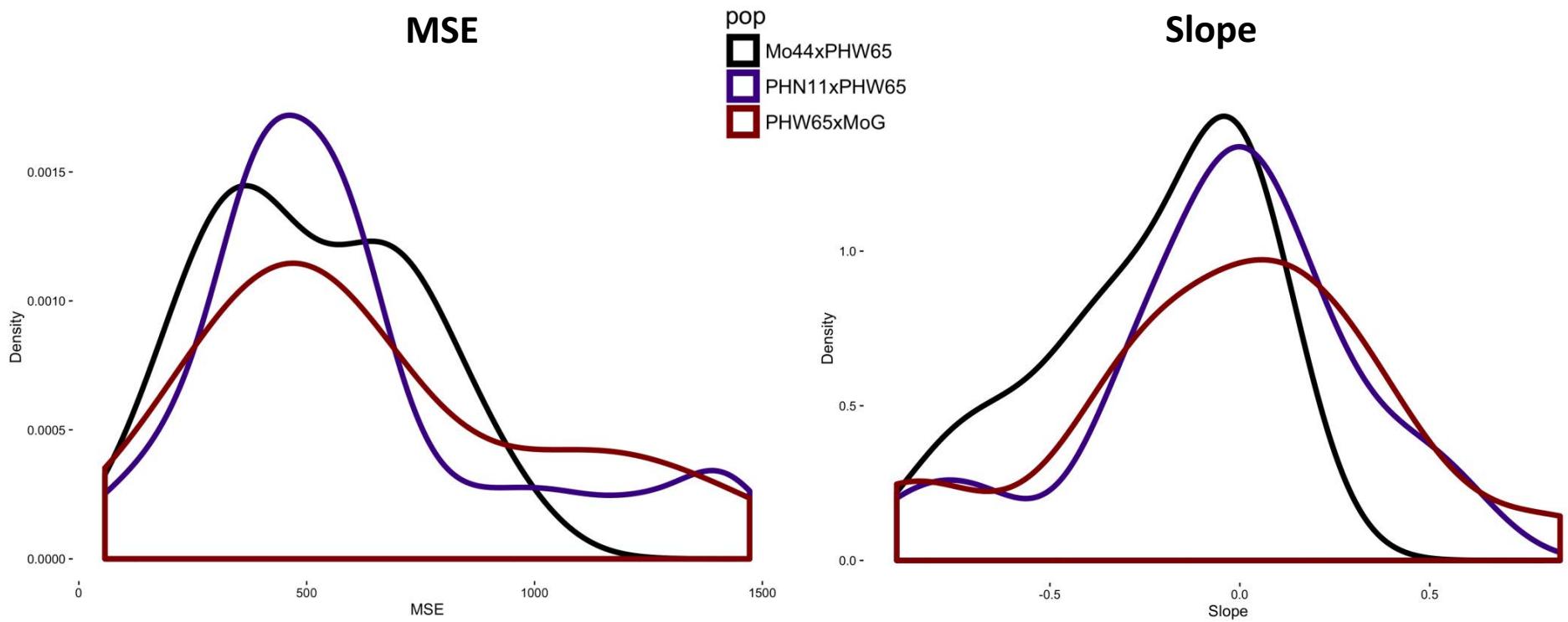
✧ Naser Alkhalifah (UW)
✧ Martin Bohn (UIUC)
✧ Ed Buckler (ARS)
✧ Darwin Campbell (ISU)
✧ Ignacio Ciampitti (KSU)
✧ James Clohessy (Cornell)
✧ Liang Dong (ISU)
✧ Jode Edwards (ARS)
✧ David Ertl (IA Corn)
✧ Sherry Flint-Garcia (ARS)
✧ Joseph Gage (UW)
✧ Jack Gardiner (ISU)
✧ Fiona Goggin (Univ AR)
✧ Byron Good (Guelph)
✧ Mike Gore (Cornell)
✧ Christopher Graham (SDSU)
✧ Patricio Grassini (UNL)
✧ Jerry Hatfield (ARS)
✧ Brien Henry (MSU)
✧ Candy Hirsch (UMN)
✧ Jim Holland (ARS)
✧ Elizabeth Hood (AR-State)
✧ David Hooker (Guelph)

✧ Diego Jarquin (UNL)
✧ Shawn Kaepller (UW)
✧ Joe Knoll (ARS)
✧ Judith Kolkman (Cornell)
✧ Greg Kruger (UNL)
✧ Nick Lauter (ARS)
✧ Carolyn Lawrence-Dill (ISU)
✧ Liz Lee (Guelph)
✧ Zhizhai Liu (TAMU)
✧ Natalia de Leon (UW)
✧ Alex Lipka (UIUC)
✧ Argelia Lorence (AR-State)
✧ Aaron Lorenz (UMN)
✧ Jonathan Lynch (PSU)
✧ John McKay (CSU)
✧ Nathan Miller (UW)
✧ Steve Moose (UIUC)
✧ Seth Murray (TAMU)*
✧ Rebecca Nelson (Cornell)
✧ Torbert Rochedorf (Purdue)
✧ Oscar Rodriguez (UNL)
✧ Cinta Romay (Cornell)
✧ Emily Rothfusz (UW)

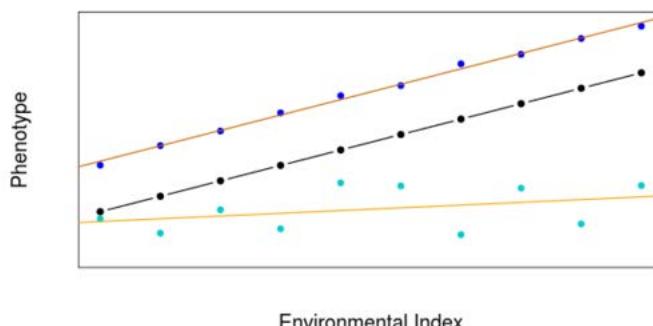
✧ James Schnable (UNL)
✧ Pat Schnable (ISU)
✧ Brian Scully (ARS)
✧ Rajandep Sekhon (Clemson)
✧ Maninder Singh (MSU)
✧ Kevin Silverstein (UMN)
✧ Margaret Smith (Cornell)
✧ Bob Snyder (PSU)
✧ Edgar Spalding (UW)
✧ Nathan Springer (UMN)
✧ Srikant Srinivasan (ISU)
✧ Yiwei Sun (ISU)
✧ Kurt Thelen (MSU)
✧ Peter Thomison (OSU)
✧ Kelly Thorp (ARS)
✧ Mitch Tuinstra (Purdue)
✧ Jason Wallace (UGA)
✧ Renee Walton (ISU)
✧ Rick Ward (UA)
✧ Bill Widdicombe (MSU)
✧ Rod Williamson (IA Corn)
✧ Randy Wisser (UDel)
✧ Wenwei Xu (TAMU)
✧ Cheng-Ting Yeh (ISU)
✧ Jianming Yu (ISU)



2014/15 Summary – Hybrids Span Range of Slope and MSE Across Testers – Grain Yield:



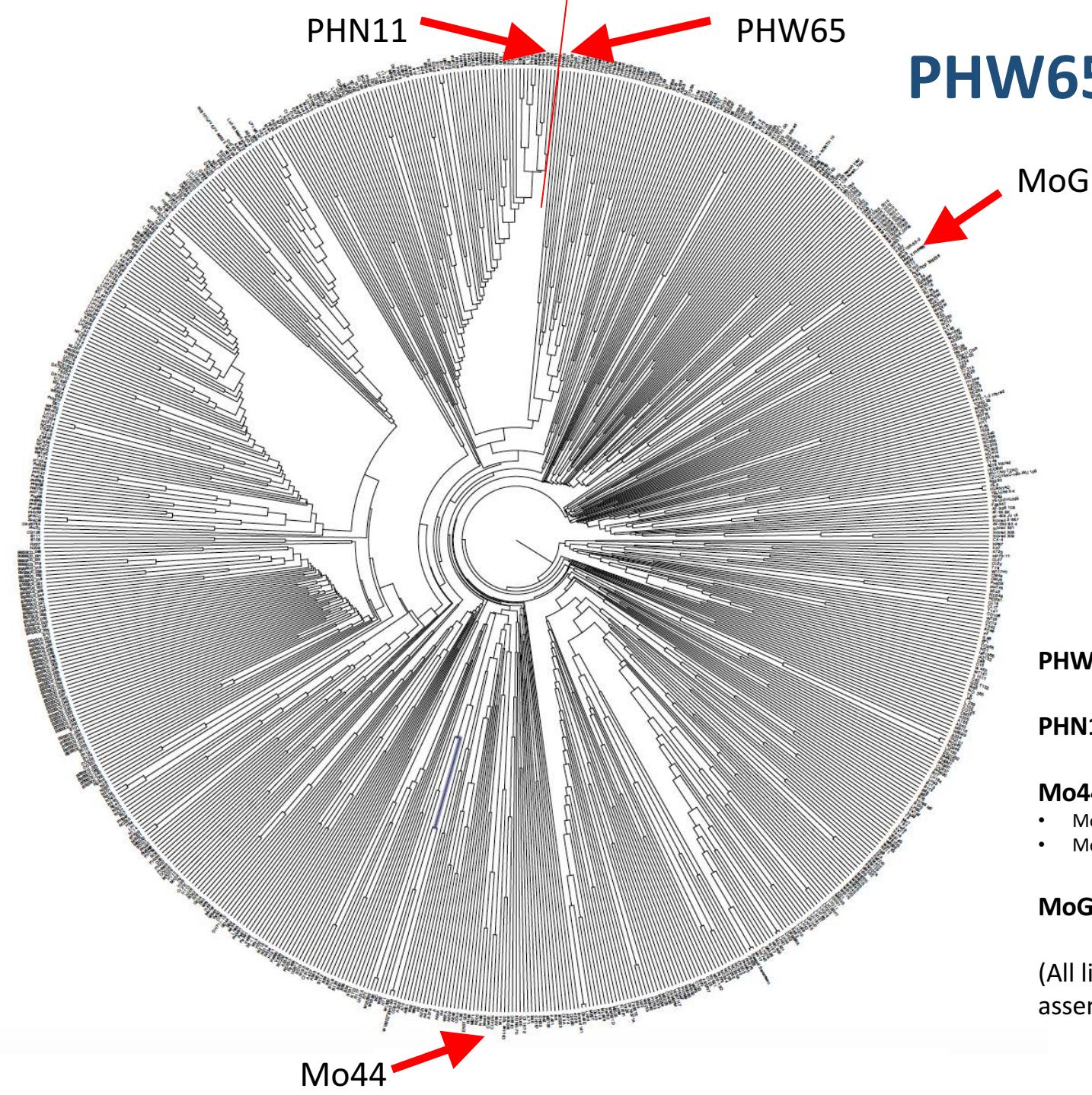
❖ 6 to 8 location per hybrid



2016 Testcross Data: 313 lines by PHB47

	Yield (bu/A)			Moist (%)		
	AVE	MAX	MIN	AVE	MAX	MIN
PHN11 X PHW65	183.4	135.8	208.5	17.8	16.2	19.4
Mo44 X PHW65	177.7	134.6	204.0	18.1	16.1	19.6
PHW65 X MoG	166.1	134.6	193.6	17.8	16.1	19.3

PHW65 Populations



PHW65: PH861 X PH595

PHN11: PH806 X PH207(2)

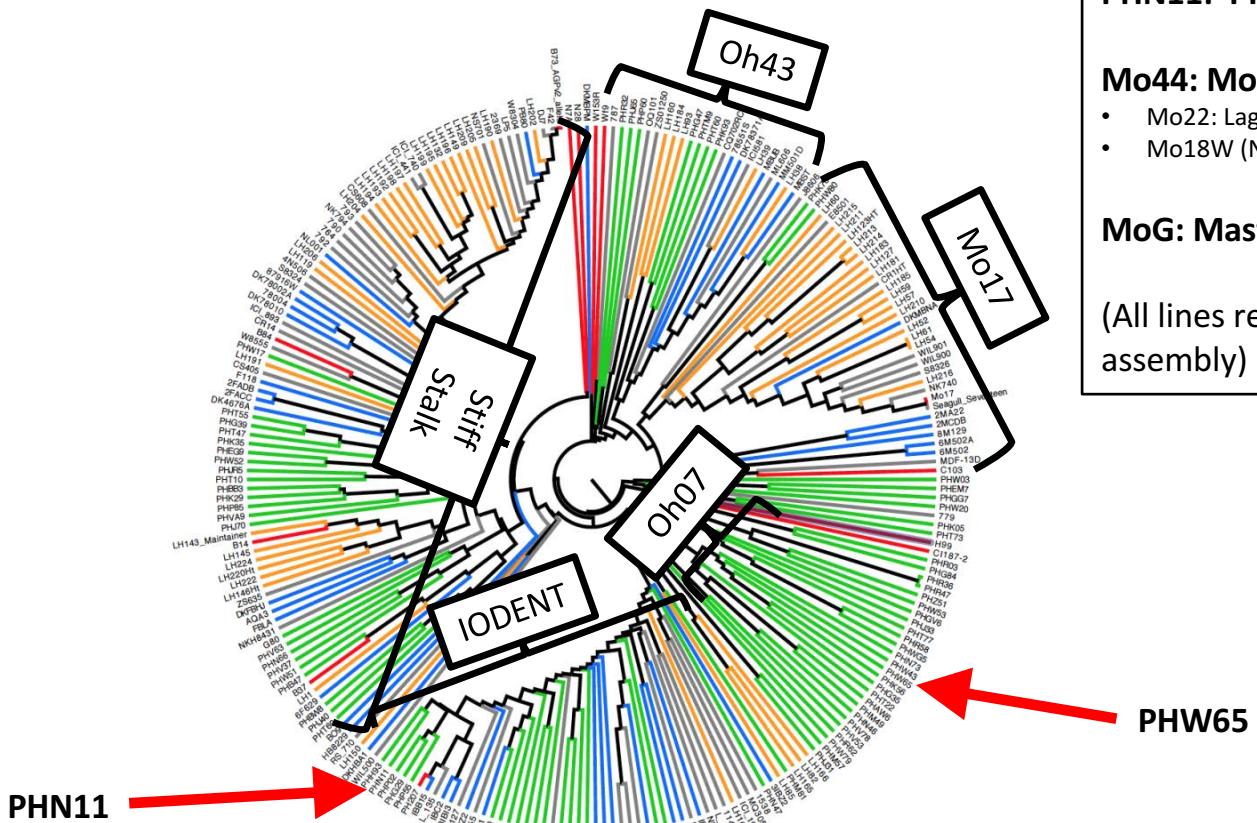
Mo44: Mo22 X Pioneer Mex Syn 17

- Mo22: Laguna
- Mo18W (NAM Parent): Wf9 x Mo22(2)

MoG: Mastadon

(All lines resequenced; PH207 assembly)

PHW65 Populations – ex-PVPs Comparison:



PHW65: PH861 x PH595

PHN11: PH806 x PH207(2)

Mo44: Mo22 x Pioneer Mex Syn 17

- Mo22: Laguna
- Mo18W (NAM Parent): Wf9 x Mo22(2)

MoG: Mastodon

(All lines resequenced; PH207 assembly)